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***Schistosoma mansoni*-specific immune responses and allergy in Uganda**

Running title: *S. mansoni*-specific immune responses and allergy in Uganda

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Disclosures

None

Abstract

Low allergy-related disease (ARD) prevalence in low-income countries may be partly attributed to helminth infections. In the *Schistosoma mansoni* (*Sm*)-endemic Lake Victoria islands (Uganda), we recently observed positive helminth-allergy associations, despite low ARD prevalence. To understand how *Sm*-induced cytokine and antibody profiles might influence allergic response profiles in this population, we assessed *Schistosoma* worm (SWA)- and egg antigen (SEA)-specific Th1 (IFN- γ), Th2 (IL-5, IL-13) and regulatory (IL-10) cytokine profiles (n=407), and total (n=471), SWA-, SEA- and allergen (house dust mite [HDM] and cockroach)-specific (as)IgE and IgG4 profiles (n=2117) by ELISA. Wheeze was inversely associated with SWA-specific IFN- γ (p<0.001) and IL-10 (p=0.058), and SEA-specific IL-5 (p=0.004). Conversely, having a detectable asIgE response was positively associated with SWA-specific IL-5 (p=0.006) and IL-10 (p<0.001). Total, SWA-, SEA- and allergen-specific IgE and IgG4 responses were higher among *Sm* Kato-Katz positive (*Sm*KK+) and skin prick test (SPT)+ individuals compared to *Sm*KK- and SPT- individuals. However, total and asIgG4/IgE ratios were lower among SPT+ and wheezing individuals. We conclude that, in this population, helminth-induced antibody and cytokine responses may underlie individual positive helminth-atopy associations, while the overall IgG4-IgE balance may contribute to the low overall prevalence of clinical allergies in such settings.

Key words: *Schistosoma* spp, allergy, cytokine, immunoglobulin, ELISA

Introduction

Helminths have a small range of antigens that are strikingly homologous to common allergens.¹ These antigens induce immunoglobulin (Ig) E-mediated effector responses important for protection against helminth infection.^{2,3} To survive in the host, helminths modulate this atopic pathway, and this may have a bystander

protective effect against allergy-related disease (ARD).⁴ While several animal and human studies provide compelling evidence of this protection,^{5,6} others suggest that in some circumstances helminths may actually promote enhanced responses to allergens.^{7,8}

Mechanisms underlying helminth-allergy associations in low-income countries (LICs) are not fully understood. Hypothesised pathways that underpin these associations are shown in Figure 1. Helminth-induced cytokine and antibody profiles may influence allergic responses, and consequently epidemiological trends pertaining to ARDs.^{5,9} Both helminth- and allergen-specific immune responses are characterised by elevated Th2-type responses (interleukin [IL]-4, IL-5 and IL-13).^{10,11} Helminths, unlike allergens, further induce strong immunoregulation epitomised by IL-10 production.¹² Typically, these cytokines influence the profile of antibodies involved in helminth infection and allergy. Helminth-induced IL-10 may drive immunoglobulin class switching to IgG4^{13,14} which, akin to the Th2 cytokine-induced¹⁵ polyclonally stimulated IgE, may inhibit development of allergen-specific effector responses,^{5,16} leading to inverse helminth-allergy associations. Conversely, helminth-induced protein-specific IgE may promote strong, cross-reactive helminth- and allergen-specific responses, resulting in positive helminth-allergy associations.^{17,18}

Emerging epidemiological data on helminth-allergy associations in Uganda reflect the complex interaction between helminths and allergens: while observational analyses in a birth cohort suggested a protective effect of childhood and maternal helminths against childhood eczema¹⁹ that was reversed by maternal anthelmintic treatment,²⁰ we recently reported positive helminth-allergy associations in a survey conducted in the *Schistosoma mansoni* (*Sm*)-endemic Lake Victoria islands, albeit against a backdrop of low ARD prevalence.²¹ To establish how *Sm*-induced cytokine and antibody profiles underpinned helminth-allergy associations in the above survey, we here describe an assessment of *Sm*-specific cytokine profiles, as well as total, allergen- and *Sm*-specific IgE and IgG4 profiles, and their relationship with *Sm* infection status, wheeze and atopy.

Methods

Study population

Samples were collected during the baseline household survey preceding a cluster-randomised trial of standard versus intensive anthelmintic intervention (the Lake Victoria Island Intervention Study on Worms and Allergy-related diseases, LaVIISWA; ISRCTN47196031) described elsewhere.^{21,22} Briefly, each consenting LaVIISWA participant completed a questionnaire, provided blood, urine and stool and underwent skin prick testing (SPT). Primary allergy-related outcomes were reported wheeze in the previous 12 months and atopy. Wheeze is widely used as a surrogate for asthma in epidemiological studies,²³ and was assessed for all ages using the International Study of Asthma and Allergies in Childhood (ISAAC) questionnaire. Such symptom questionnaires have been identified as the best way to estimate asthma prevalence in epidemiological studies.²³ The ISAAC questionnaire was used to ask participants (or their caregivers) if they had ever wheezed and if so, if they had wheezed in the last 12 months. Details on aetiology were not collected. Atopy was defined as **1)** SPT reactivity to any of *Dermatophagoides* mix, *Blomia tropicalis* or German cockroach (*Blattella germanica*) [ALK-Abelló; supplied by Laboratory Specialities Ltd., South Africa], and **2)** detectable IgE response (>312ng/ml by ELISA²¹) to *Dermatophagoides pteronyssinus* [hereinafter 'house dust mite (HDM)'] and/or German cockroach whole allergen extracts (Greer Labs, U.S.A).

Ethics committees of Uganda Virus Research Institute, London School of Hygiene and Tropical Medicine and Uganda National Council for Science and Technology approved the study.

Laboratory methods

Two slides from one stool sample per individual were independently examined by different technicians for *Sm* eggs using the Kato-Katz method.²⁴

We assessed IFN- γ (Th1-type), IL-5, IL-13 (Th2-type) and IL-10 (regulatory) levels by ELISA using supernatants from six-day whole blood cultures stimulated with *Schistosoma* worm (SWA) and egg antigens (SEA), as previously described.²⁵ Briefly, heparinised blood was diluted with RPMI 1640 medium (Life technologies, UK) supplemented with penicillin, streptomycin, glutamine and Hepes buffer (all from

Life technologies, UK), plated in 96-well culture plates and stimulated (at 37°C, 5% CO₂) with 10µg/ml SWA or SEA (provided by Professor Mike Doenhoff, University of Nottingham) or mitogen (phytohaemagglutinin, PHA, Sigma, UK), or left unstimulated. Supernatants were harvested on day six and stored at -80°C until analysis. Cytokine levels in supernatants were measured by ELISA (Becton Dickinson, USA). The net response to each stimulus was calculated by subtracting the concentration in the unstimulated control well. Response values that were below the dynamic range of the assay and those that were negative after subtraction of the response in the unstimulated well were assigned a value of zero.

HDM and cockroach extract-specific IgE and IgG4 were measured in plasma using an in-house ELISA described previously.²⁰ Briefly, MICROLON® 96-well plates (Greiner bio-one, UK) were coated overnight at 4°C with 5µg/ml HDM or cockroach allergens and two-fold dilutions of human IgE (Calbiochem, Beeston, UK) or IgG4 (Sigma-Aldrich) standards. Plates were blocked at room temperature (RT) with 1% skimmed milk and incubated overnight at 4°C with plasma samples diluted 1/20 (IgE assay) or 1/40 (IgG4 assay) with 10% fetal bovine serum in PBS-Tween 20. Specific IgE or IgG4 was detected using biotinylated monoclonal mouse anti-human IgE or IgG4 (BD Pharmingen™) and a streptavidin-Horseradish Peroxidase conjugate (Mast Group Ltd, Bootle, UK). O-phenylenediamine (Sigma-Aldrich) was used as a substrate, and the reaction stopped with 2M Sulphuric acid. Optical density values were measured at 490nm (reference wavelength 630nm) on a 96-well plate ELISA reader. IgE or IgG4 concentrations (ng/ml) were interpolated from standard curves using a five-parameter curve fit using Gen5 data collection and analysis software (BioTek Instruments Inc, Vermont, Winooski, USA).

Total, SWA- and SEA-specific IgE and IgG4 ELISAs were performed using similar in-house procedures (detailed in this article's supporting information).

Statistical methods

Our hypothesised mode of action of *S. mansoni*-induced cytokines and antibodies on allergy-related outcomes is illustrated in Figure 1. Using STATA 13.1 (College Station, Texas, USA), we performed cross-sectional analyses to assess whether *Sm* Kato-Katz positivity and allergy-related outcomes were associated with antibody and cytokine levels, using the “svy” command to allow for the non-self-weighting cluster

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survey design. Raw cytokine and antibody responses were skewed, so $\log_{10}$ (concentration+1)-transformed antibody and cytokine data were used in our regression models; we back-transformed the results to obtain geometric mean ratios and 95% confidence intervals. Crude and age- and sex-adjusted analyses were performed. Associations between antibody responses were estimated using Spearman's correlation coefficient (r_s). We used a 5% significance level for all analyses. P values quoted in the main text are from adjusted analyses.

Results

Questionnaire data were obtained from 2316 participants.²² Their characteristics and those of participants for whom cytokine and antibody responses were assessed are shown in Table 1. Participants for whom cytokine (n=407) and total antibody levels (n=471) were assessed were a subset of participants who had allergen-, SWA- and SEA-specific antibody results (n=2117). Cytokine responses were assessed using samples from 1-17 year olds, to allow comparison with related cellular immunology studies in an urban birth cohort (data not shown). Allergen-, SWA- and SEA-specific responses were assessed in all survey participants that had sufficient plasma sample stored.

S. mansonii-specific cytokines and allergy-related outcomes

Individuals who tested positive for *Sm* by Kato-Katz (*Sm*KK+) had higher geometric mean concentrations of SWA-specific type 2 and regulatory cytokines compared to *Sm*KK- individuals (**Table 2**), but this was statistically significant only for IL-5 ($p=0.034$). However, there was no dose-response relationship with infection intensity (supplementary Table S3A). SEA-specific responses were similar between *Sm*KK+ and *Sm*KK- individuals.

Wheeze was inversely associated with SWA-specific IFN- γ ($p<0.001$) and IL-10 ($p=0.058$; **Table 2**), and SWA-specific IL-10/IL-5 and SEA-specific IL-10/IFN- γ ratios (supplementary **Table S1**). Wheezing individuals also had lower mean SEA-specific cytokine responses, although statistical evidence for an inverse association was observed only for SEA-specific IL-5 ($p=0.004$).

Conversely, there was a crude positive association between SPT positivity and SWA-specific IL-10 ($p=0.048$; Table 2); and individuals with detectable allergen-specific (as)IgE had higher SWA-specific IL-5 ($p=0.006$) and IL-10 responses ($p<0.001$; Table 2) and higher SWA- and SEA-specific IL-10/IFN- γ and IL-5/IFN- γ ratios (Table S1).

Antibody responses, S. mansoni Kato-Katz positivity and allergy

Kato-Katz positivity was strongly positively associated with SWA- and SEA-specific IgE and IgG4 ($p<0.001$), with HDM-specific IgE ($p=0.006$) and HDM- and cockroach-specific IgG4 ($p<0.001$; Table 3 and supplementary Figure S1). However, correlations between *Sm* antigen-specific antibodies and allergen-specific antibodies were weak ($r_s<0.4$, Table S2).

Kato-Katz positivity was also strongly positively associated with total IgE ($p<0.001$) (Table 3 and Figure S1), which was in turn weakly correlated with SEA-specific IgE but moderately correlated with SWA-specific IgE ($r_s=0.31$ and $r_s=0.51$, respectively; Table S2). Similarly, total IgG4 ($p<0.001$), total IgG4/total IgE ratios ($p=0.005$) and total IgE/asIgE ratios ($p<0.05$) were positively associated with Kato-Katz positivity. In addition, there was a general dose-response relationship between *S. mansoni* infection intensity and antibody responses (supplementary Table S3B).

Cockroach-specific IgE and total IgE were positively associated with cockroach SPT reactivity. HDM-specific IgE and IgG4, SWA- and SEA-specific IgE and total IgE, were all positively associated with HDM SPT reactivity (Table 3 and Figure S1). In contrast, cockroach SPT reactivity was inversely associated with total IgG4/total IgE ratios ($p=0.022$), and HDM SPT reactivity with total IgE/HDM-specific IgE ratios ($p<0.001$).

Associations between wheeze and antibody responses (Table 3 and Figure S1), when significant, were inverse. HDM IgG4/IgE ratios ($p=0.032$), cockroach-specific IgE ($p=0.003$) and cockroach-specific IgG4 ($p=0.001$) were all inversely associated with wheeze.

Discussion

In this highly-*Sm*-endemic setting, associations between wheeze and *Sm*-specific cytokines and antibodies, when significant, were inverse. However, SPT reactivity and detectable asIgE were positively associated with the same *Sm*-specific responses.

In this population, *Sm* exposure is almost universal, and infection much higher than indicated by Kato-Katz: urine assessment for *Sm* circulating cathodic antigen (CCA) indicated a prevalence of over 70%, compared to 51.4% prevalence by Kato-Katz.²² Therefore, Kato-Katz negativity in many study participants was indicative of lighter (rather than absent) infection. This explains why, although SWA-specific Th2-type and regulatory cytokine responses were generally higher among SmKK+ individuals, only SWA-specific IL-5 reached significant levels, and why SEA-specific responses were similar between SmKK+ and SmKK- individuals. Further support for these observations comes from supplementary analysis (Table S4A), which shows that cytokine responses were similar between SmKK-CCA+ and SmKK+CCA+/- individuals.

All statistically significant associations between atopy and *Sm*-specific cytokine responses were positive. Associations with whole blood cytokine responses are best interpreted taking into account total cell counts, but these data were unavailable. However, atopy-antibody associations were also positive. Besides, these results mirror our previous epidemiological observations in this population, where *Sm* infection was positively associated with *Dermatophagoides*-specific IgE, and atopy-wheeze associations were stronger in presence of *Sm* infection.²¹

Our results were unexpected in view of earlier findings from Gabon⁹ which showed an inverse association between dust mite SPT and SWA-specific IL-10 (albeit we used whole blood cultures, compared to peripheral blood mononuclear cells in the Gabon study). However, although IL-10 is chiefly immunomodulatory,²⁶⁻²⁸ it may also enhance IgE production in already IgE-switched B cells;¹³ these may be abundant in individuals from this helminth-endemic setting. SWA- and SEA-specific IgE were weakly positively associated with HDM SPT reactivity, perhaps unsurprisingly, as helminth antigens may induce cross-reactive helminth-and allergen-specific IgE effector responses. Total serum IgE, elevated during helminth infection mainly due to increased synthesis of polyclonal IgE, has been proposed to inhibit allergic responses.^{29,30} However, contrasting evidence links high serum IgE levels to

increased expression of IgE receptors on human basophils,³¹ and we show positive associations between total IgE and SPT reactivity to both cockroach and dust mite.

In keeping with the original hypothesis, associations between wheeze and cytokine and antibody responses, when significant, were inverse. Furthermore, total and allergen-specific IgG4/IgE ratios were mostly inversely associated with atopy, implying that the regulatory role of IgG4 against allergy might best be assessed relative to IgE. Also, lower total/asIgE ratios among HDM SPT+ individuals are consistent with the perception that high total/asIgE ratios may be protective against allergic responses, because non-specific polyclonal IgE may compete with asIgE to saturate IgE receptors.²⁹

One limitation of assessing helminth-allergy associations and underlying mechanisms in this population is the almost universal exposure to helminths, and lack of data on duration of infection. We also report a large number of statistical tests, so some apparently “significant” findings could have occurred by chance. Since we anticipated that some of our measures might be correlated, we did not formally adjust for multiplicity, instead we focussed on patterns of association and consistency of results, and on biological plausibility with reference to other findings. Another potential limitation is that wheeze was relatively rare in the study population and hence some of our comparison groups (such as the age group 1-17 years) had a low prevalence. Besides, reported wheeze could easily be misclassified in this population due to lack of a direct translation of ‘wheeze’ in the native languages.²¹

Nonetheless, our results generally agree with our epidemiological observations in the same population,²¹ where we found a very low prevalence of clinical allergies, despite positive helminth-atopy associations.

LaVIISWA trial team

Project leaders, physicians, post-doctoral scientists: Richard Sanya, Margaret Nampijja, Harriet Mpairwe, Geraldine O’Hara, Barbara Nerima. **Laboratory staff and collaborators:** Gyaviira Nkurunungi, Joyce Kabagenyi, Dennison Kizito, John Vianney Tushabe, Jacent Nassuuna, Jaco Verweij, Stephen Cose, Linda Wammes, Prossy Kabuubi, Emmanuel Niwagaba, Gloria Oduru, Grace Kabami, Elson Abayo, Eric Ssebagala, Fred Muwonge. **Statisticians and data managers:** Emily Webb, Remy Hoek Spaans, Lawrence Muhangi, Lawrence Lubyayi, Helen Akurut, Fatuma

Nalukenge, Beatrice Mirembe, Justin Okello, Sebastian Owilla, Jonathan Levin, Stephen Nash. **Clinical officers:** Milly Namutebi, Christopher Zziwa. **Nurses:** Esther Nakazibwe, Josephine Tumusiime, Caroline Ninsiima, Susan Amongi, Grace Kamukama, Susan Iwala, Florence. **Internal monitor:** Mirriam Akello. **Field workers:** Robert Kizindo, Moses Sewankambo, Denis Nsubuga. **Social sciences:** Edward Tumwesige. **Boatman:** David Abiriga. **Driver:** Richard Walusimbi. **HIV counselling and testing:** Victoria Nannozi, Cynthia Kabonesa. **Vector Control Programme staff:** James Kaweesa, Edridah Tukahebwa. **Administrative management:** Moses Kizza. **Principal investigator:** Alison Elliott.

Author contributions

AME conceived the main study. AME, RES and MN led the field and clinic teams. AME, GN and JK participated in the design of laboratory studies. GN, JK, BW and JN performed the experiments. GN and ELW analysed the results. GN wrote the manuscript, with all authors contributing to the interpretation of the results, and revision and approval of the final manuscript. GN is the guarantor of the paper.

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References

1. Tyagi N, Farnell EJ, Fitzsimmons CM, et al. Comparisons of Allergenic and Metazoan Parasite Proteins: Allergy the Price of Immunity. *PLoS Comput Biol* 2015; **11**(10): e1004546.
2. Dunne DW, Butterworth AE, Fulford AJ, et al. Immunity after treatment of human schistosomiasis: association between IgE antibodies to adult worm antigens and resistance to reinfection. *Eur J Immunol* 1992; **22**(6): 1483-94.
3. Dunne DW, Webster M, Smith P, et al. The isolation of a 22 kDa band after SDS-PAGE of *Schistosoma mansoni* adult worms and its use to demonstrate that IgE responses against the antigen(s) it contains are associated with human resistance to reinfection. *Parasite Immunol* 1997; **19**(2): 79-89.
4. Smits HH, Everts B, Hartgers FC, Yazdanbakhsh M. Chronic helminth infections protect against allergic diseases by active regulatory processes. *Curr Allergy Asthma Rep* 2010; **10**(1): 3-12.
5. Hamid F, Amoah AS, van Ree R, Yazdanbakhsh M. Helminth-induced IgE and protection against allergic disorders. *Curr Top Microbiol Immunol* 2015; **388**: 91-108.
6. Maizels RM, McSorley HJ, Smyth DJ. Helminths in the hygiene hypothesis: sooner or later? *Clin Exp Immunol* 2014; **177**(1): 38-46.
7. Santiago Hda C, Ribeiro-Gomes FL, Bennuru S, Nutman TB. Helminth infection alters IgE responses to allergens structurally related to parasite proteins. *J Immunol* 2015; **194**(1): 93-100.
8. Levin M, Muloiwa R, Le Souef P, Motala C. Ascaris sensitization is associated with aeroallergen sensitization and airway hyperresponsiveness but not allergic disease in urban Africa. *J Allergy Clin Immunol* 2012; **130**(1): 265-7.
9. van den Biggelaar AH, van Ree R, Rodrigues LC, et al. Decreased atopy in children infected with *Schistosoma haematobium*: a role for parasite-induced interleukin-10. *Lancet* 2000; **356**(9243): 1723-7.
10. Allen JE, Maizels RM. Diversity and dialogue in immunity to helminths. *Nat Rev Immunol* 2011; **11**(6): 375-88.

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11. Licona-Limon P, Kim LK, Palm NW, Flavell RA. TH2, allergy and group 2 innate lymphoid cells. *Nat Immunol* 2013; **14**(6): 536-42.
 12. King CL, Medhat A, Malhotra I, et al. Cytokine control of parasite-specific anergy in human urinary schistosomiasis. IL-10 modulates lymphocyte reactivity. *J Immunol* 1996; **156**(12): 4715-21.
 13. Jeannin P, Lecoanet S, Delneste Y, Gauchat JF, Bonnefoy JY. IgE versus IgG4 production can be differentially regulated by IL-10. *J Immunol* 1998; **160**(7): 3555-61.
 14. Tsuboi H, Matsuo N, Iizuka M, et al. Analysis of IgG4 class switch-related molecules in IgG4-related disease. *Arthritis Res Ther* 2012; **14**(4): R171.
 15. Poulsen LK, Hummelshoj L. Triggers of IgE class switching and allergy development. *Ann Med* 2007; **39**(6): 440-56.
 16. James LK, Till SJ. Potential Mechanisms for IgG4 Inhibition of Immediate Hypersensitivity Reactions. *Curr Allergy Asthma Rep* 2016; **16**(3): 23.
 17. Santiago HC, Bennuru S, Boyd A, Eberhard M, Nutman TB. Structural and immunologic cross-reactivity among filarial and mite tropomyosin: implications for the hygiene hypothesis. *J Allergy Clin Immunol* 2011; **127**(2): 479-86.
 18. Santiago HC, LeeVan E, Bennuru S, et al. Molecular mimicry between cockroach and helminth glutathione S-transferases promotes cross-reactivity and cross-sensitization. *J Allergy Clin Immunol* 2012; **130**(1): 248-56 e9.
 19. Mpairwe H, Ndibazza J, Webb EL, et al. Maternal hookworm modifies risk factors for childhood eczema: results from a birth cohort in Uganda. *Pediatr Allergy Immunol* 2014; **25**(5): 481-8.
 20. Mpairwe H, Webb EL, Muhangi L, et al. Anthelmintic treatment during pregnancy is associated with increased risk of infantile eczema: randomised-controlled trial results. *Pediatr Allergy Immunol* 2011; **22**(3): 305-12.
 21. Webb EL, Nampijja M, Kaweesa J, et al. Helminths are positively associated with atopy and wheeze in Ugandan fishing communities: results from a cross-sectional survey. *Allergy* 2016; **71**(8): 1156-69.

22. Nampijja M, Webb EL, Kaweesa J, et al. The Lake Victoria island intervention study on worms and allergy-related diseases (LaVIISWA): study protocol for a randomised controlled trial. *Trials* 2015; **16**(1): 187.
23. Pekkanen J, Pearce N. Defining asthma in epidemiological studies. *Eur Respir J* 1999; **14**(4): 951-7.
24. Katz N, Chaves A, Pellegrino J. A simple device for quantitative stool thick-smear technique in *Schistosomiasis mansoni*. *Rev Inst Med Trop Sao Paulo* 1972; **14**(6): 397-400.
25. Elliott AM, Mawa PA, Webb EL, et al. Effects of maternal and infant co-infections, and of maternal immunisation, on the infant response to BCG and tetanus immunisation. *Vaccine* 2010; **29**(2): 247-55.
26. de Waal Malefyt R, Yssel H, de Vries JE. Direct effects of IL-10 on subsets of human CD4+ T cell clones and resting T cells. Specific inhibition of IL-2 production and proliferation. *J Immunol* 1993; **150**(11): 4754-65.
27. Ding L, Linsley PS, Huang LY, Germain RN, Shevach EM. IL-10 inhibits macrophage costimulatory activity by selectively inhibiting the up-regulation of B7 expression. *J Immunol* 1993; **151**(3): 1224-34.
28. Ding L, Shevach EM. IL-10 inhibits mitogen-induced T cell proliferation by selectively inhibiting macrophage costimulatory function. *J Immunol* 1992; **148**(10): 3133-9.
29. Godfrey RC, Gradidge CF. Allergic sensitisation of human lung fragments prevented by saturation of IgE binding sites. *Nature* 1976; **259**(5543): 484-6.
30. Lynch NR, Hagel IA, Palenque ME, et al. Relationship between helminthic infection and IgE response in atopic and nonatopic children in a tropical environment. *J Allergy Clin Immunol* 1998; **101**(2 Pt 1): 217-21.
31. MacGlashan DW, Jr., Bochner BS, Adelman DC, et al. Down-regulation of Fc(epsilon)RI expression on human basophils during in vivo treatment of atopic patients with anti-IgE antibody. *J Immunol* 1997; **158**(3): 1438-45.

Legends to figures

Figure 1.

We hypothesise that the Th2 cytokine-induced *Sm*-specific IgE promotes potent, *Sm*-specific, atopic effector responses and *Sm* elimination from the host, and also cross-reactive responses to some allergens, resulting in positive *Sm*-allergy associations. By contrast, *Sm*-specific IL-10, IgG4 and/or non-specific polyclonally stimulated IgE inhibit these allergy-related outcomes. White and shaded arrows denote promotion and inhibition, respectively.

Table 1. Characteristics of participants

Characteristic	Survey population (N=2316), n/N (%)	Immunological measure, n/N (%)		
		Cytokine responses [‡] (N=407)	Allergen-, SWA- and SEA- specific IgE and IgG4 [§] (N=2117)	Total IgE and IgG4 [¶] (N=471)
Age in years, median (IQR)	24 (8, 32)	9 (6, 16)	25 (10, 33)	19.5 (3, 31.25)
Male sex	1268/2316 (54.7)	168/407 (41.3)	1152/2117 (54.4)	225/471 (47.7)
PZQ in last 12 months	382/2255 (16.9)	48/393 (12.2)	368/2062 (17.8)	15/459 (15.5)
Helminth infections				
<i>S. mansoni</i> (KK)	1041/1996 (51.4)	204/373 (54.7)	1008/1882 (53.6)	184/428 (42.9)
<i>S. mansoni</i> (urine CCA)	661/917 (72.0)	94/128 (73.4)	634/875 (72.5)	101/152 (66.5)
<i>S. mansoni</i> intensity (KK)				
Uninfected	995/1996 (48.6)	169/373 (45.3)	874/1882 (46.4)	244/428 (57.0)
Low	429/1996 (21.0)	77/373 (20.6)	411/1882 (21.8)	70/428 (16.4)
Moderate	288/1996 (13.7)	56/373 (15.0)	279/1882 (14.8)	51/428 (11.9)
Heavy	324/1996 (16.6)	71/373 (19.0)	318/1882 (16.9)	63/428 (14.7)
Any nematode infection [†]	788/2004 (39.3)	129/373 (34.6)	738/1889 (39.1)	87/428 (20.3)
Allergy-related outcomes				
Wheeze in last 12 months	107/2301 (4.7)	14/404 (3.5)	106/2103 (5.04)	58/468 (12.4)
Atopy (SPT)				
Any	404/1976 (19.1)	78/372 (20.9)	403/1961 (20.6)	135/448 (30.1)
<i>Dermatophagoides</i>	190/1978 (9.0)	33/372 (8.9)	189/1963 (9.6)	61/448 (13.6)

<i>Blomia</i>	205/1976 (9.6)	31/372 (8.3)	204/1961 (10.4)	67/447 (14.9)
Cockroach	272/1977 (13.2)	61/372 (16.4)	272/1962 (13.9)	90/448 (20.1)
Atopy (detectable asIgE)				
Any	1685/2117 (79.6)	320/403 (79.4)	1685/2117 (79.6)	358/471 (76.0)
<i>Dermatophagoides</i>	1534/2115 (72.5)	278/403 (68.9)	1534/2115 (72.5)	326/471 (69.2)
Cockroach	886/2117 (41.9)	183/403 (45.4)	886/2117 (41.9)	186/471 (39.5)

[‡]Assessed using samples from 1-17 year olds, to allow comparison with related cellular immunology studies in an urban birth cohort (data not shown here).

[§]Assessed in all survey participants that had sufficient plasma sample stored.

[¶]Samples randomly selected from individuals with antigen-specific antibody data.

[†]Infection with any of *Ascaris lumbricoides*, *Trichuris trichiura* (assessed by KK), *Necator americanus*, *Strongyloides stercoralis* (assessed by PCR) and *Mansonella perstans* (assessed by modified Knott's method).

PZQ: Praziquantel treatment; **KK:** Kato-Katz; **CCA:** circulating cathodic antigen; **SPT:** skin prick test; **SWA:** *Schistosoma* worm antigen; **SEA:** *Schistosoma* egg antigen; **asIgE:** allergen-specific IgE

Table 2. Associations between *S. mansoni*-specific cytokine levels and i) *S. mansoni* infection status, ii) reported wheeze and iii) atopy (SPT reactivity and detectable allergen-specific IgE)

Antigen	Cytokine	Geometric mean [†]		Unadjusted		Adjusted for age and sex	
				GMR (95% CI) [‡]	p value	GMR (95% CI) [‡]	p value
		SmKK- n=169	SmKK+ n=204				
SWA	IFN- γ	1.16	1.13	1.06 (0.86, 1.30)	0.542	1.05 (0.87, 1.28)	0.531
	IL-5	14.92	49.47	1.43 (1.13, 1.81)	0.005	1.32 (1.02, 1.71)	0.034
	IL-13	7.01	17.56	1.20 (0.94, 1.54)	0.132	1.15 (0.88, 1.48)	0.282
	IL-10	3.99	11.58	1.21 (0.97, 1.51)	0.084	1.16 (0.91, 1.48)	0.207
SEA	IFN- γ	0.73	0.56	0.97 (0.81, 1.17)	0.760	0.98 (0.82, 1.18)	0.884
	IL-5	5.02	3.11	0.84 (0.59, 1.19)	0.320	0.84 (0.58, 1.19)	0.319
	IL-13	2.25	1.95	0.86 (0.71, 1.05)	0.127	0.88 (0.73, 1.06)	0.190
	IL-10	3.19	4.42	0.93 (0.78, 1.13)	0.486	0.93 (0.76, 1.13)	0.477
		No wheeze n=390	Wheeze n=14				
SWA	IFN- γ	1.27	0.23	0.60 (0.45, 0.80)	0.001	0.57 (0.44, 0.76)	<0.001
	IL-5	29.12	27.59	1.29 (0.72, 2.33)	0.373	1.14 (0.63, 2.08)	0.657
	IL-13	11.75	9.78	1.26 (0.65, 2.45)	0.465	1.17 (0.58, 2.36)	0.635
	IL-10	7.91	2.03	0.69 (0.47, 1.01)	0.059	0.66 (0.43, 1.02)	0.058
SEA	IFN- γ	0.66	0.33	0.83 (0.66, 1.04)	0.101	0.83 (0.65, 1.05)	0.121
	IL-5	4.37	1.20	0.52 (0.33, 0.83)	0.007	0.51 (0.33, 0.79)	0.004
	IL-13	2.25	0.96	0.75 (0.44, 1.25)	0.256	0.76 (0.44, 1.33)	0.327
	IL-10	4.34	0.59	0.70 (0.35, 1.38)	0.295	0.71 (0.37, 1.36)	0.291
		SPT- n=294	SPT+ [§] n=78				
SWA	IFN- γ	1.03	1.73	1.15 (0.96, 1.36)	0.115	1.13 (0.94, 1.34)	0.178
	IL-5	29.23	39.73	1.15 (0.86, 1.52)	0.330	1.02 (0.75, 1.38)	0.897
	IL-13	13.54	10.32	0.99 (0.72, 1.36)	0.961	0.92 (0.67, 1.26)	0.596

SEA	IL-10	6.79	12.00	1.25 (1.00, 1.55)	0.048	1.21 (0.95, 1.54)	0.126
	IFN- γ	0.55	0.99	1.11 (0.82, 1.51)	0.493	1.13 (0.83, 1.52)	0.423
	IL-5	4.54	3.68	1.00 (0.70, 1.45)	0.965	0.98 (0.67, 1.43)	0.935
	IL-13	2.81	1.09	0.84 (0.56, 1.25)	0.376	0.85 (0.58, 1.25)	0.413
	IL-10	4.12	3.09	0.94 (0.76, 1.16)	0.560	0.96 (0.75, 1.22)	0.730
		Undetectable asIgE n=83	Detectable asIgE [¶] n=320				
SWA	IFN- γ	1.73	1.11	0.87 (0.62, 1.24)	0.444	0.86 (0.60, 1.23)	0.396
	IL-5	16.44	34.25	1.43 (1.16, 1.75)	0.001	1.32 (1.09, 1.61)	0.006
	IL-13	9.75	12.66	1.09 (0.77, 1.55)	0.599	1.04 (0.74, 1.47)	0.806
	IL-10	3.76	9.26	1.34 (1.18, 1.51)	<0.001	1.30 (1.16, 1.46)	<0.001
SEA	IFN- γ	1.12	0.56	0.79 (0.56, 1.13)	0.190	0.79 (0.57, 1.11)	0.176
	IL-5	3.78	4.17	1.13 (0.81, 1.58)	0.459	1.12 (0.82, 1.54)	0.450
	IL-13	2.21	2.21	0.88 (0.65, 1.22)	0.449	0.90 (0.67, 1.21)	0.484
	IL-10	3.49	4.17	1.05 (0.92, 1.21)	0.424	1.06 (0.93, 1.21)	0.382

[†]All cytokine concentrations in pg/ml.

[‡]Geometric mean ratios and 95% confidence intervals adjusted for the survey design.

[§]SPT reactivity to any one of *Dermatophagoides mix*, *Blomia tropicalis* or *Blattella germanica*.

[¶]Detectable IgE to either *Dermatophagoides pteronyssinus* or *Blattella germanica*.

SmKK-: Kato-Katz negative result (*S. mansoni*), single stool sample; **SmKK+**: Kato-Katz positivity for *S. mansoni*, single stool sample; **SPT**: skin prick test; **SWA**: *Schistosoma* worm antigen; **SEA**: *Schistosoma* egg antigen; **asIgE**: allergen-specific IgE; **GMR**: geometric mean ratio; **95% CI**: 95% confidence interval. P values ≤ 0.05 are highlighted in bold.

Table 3. Associations between antibody (IgE and IgG4) levels and Kato-Katz positivity (*S. mansoni*), SPT reactivity and reported wheeze

Antigen	Antibody / antibody ratio	Geometric mean [†]		aGMR (95% CI) ^{‡¶}	p value
		<i>SmKK</i> -	<i>SmKK</i> +		
SWA	IgE	1080	2433	1.54 (1.28, 1.84)	<0.001
	IgG4	4031	27355	3.71 (3.14, 4.37)	<0.001
SEA	IgE	1412	1833	1.32 (1.15, 1.52)	<0.001
	IgG4	18962	241763	5.51 (4.55, 6.67)	<0.001
House dust mite	IgE	0.782	10.678	1.25 (1.07, 1.45)	0.006
	IgG4	0.001	0.192	1.79 (1.51, 2.13)	<0.001
	IgG4/IgE ratio	0.002	0.033	1.18 (0.58, 2.41)	0.629
Cockroach	IgE	18.8	19.2	1.00 (0.82, 1.22)	0.989
	IgG4	0.002	0.292	1.50 (1.34, 1.68)	<0.001
	IgG4/IgE ratio	0.001	0.027	1.32 (0.94, 1.85)	0.110
	Total IgE	969	3073	1.37 (1.22, 1.54)	<0.001
	Total IgG4	51453	233745	1.94 (1.49, 2.52)	<0.001
	Total IgG4/ total IgE ratio	52.16	75.24	1.36 (1.11, 1.67)	0.005
	Total IgE/ cockroach IgE ratio	3.79	12.60	1.32 (1.06, 1.66)	0.014
	Total IgE/ dust mite IgE ratio	0.562	1.301	1.13 (1.02, 1.25)	0.016
		<i>Cockroach</i> <i>SPT</i> -	<i>Cockroach</i> <i>SPT</i> +		

SWA	IgE	1704	1894	1.12 (0.94, 1.32)	0.173
	IgG4	12860	14155	1.04 (0.85, 1.28)	0.675
SEA	IgE	1611	1876	1.12 (0.97, 1.29)	0.092
	IgG4	84831	101778	1.08 (0.92, 1.27)	0.319
Dust mite	IgE	2.6	42.2	1.59 (1.35, 1.89)	<0.001
	IgG4	0.022	0.061	1.06 (0.87, 1.29)	0.498
	IgG4/IgE ratio	0.009	0.001	0.56 (0.36, 0.85)	0.010
Cockroach	IgE	18.9	39.1	1.25 (1.08, 1.46)	0.004
	IgG4	0.054	0.686	1.15 (0.95, 1.39)	0.129
	IgG4/IgE ratio	0.003	0.017	0.69 (0.47, 1.02)	0.064
	Total IgE	1462	2787	1.22 (1.05, 1.42)	0.011
	Total IgG4	90643	126688	0.84 (0.65, 1.07)	0.163
	Total IgG4/ total IgE ratio	60.41	45.80	0.75 (0.58, 0.95)	0.022
	Total IgE/ cockroach IgE ratio	6.07	9.22	1.01 (0.82, 1.25)	0.894
	Total IgE/ dust mite IgE ratio	0.849	0.868	0.93 (0.84, 1.03)	0.140

		<i>Dust mite SPT-</i>	<i>Dust mite SPT+</i>		
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SWA	IgE	1667	2409	1.26 (1.00, 1.57)	0.043
	IgG4	13088	12565	1.04 (0.79, 1.36)	0.744
SEA	IgE	1623	1887	1.23 (0.99, 1.53)	0.055
	IgG4	85471	102026	1.20 (0.85, 1.68)	0.271
Dust mite	IgE	2.5	242.9	2.10 (1.57, 2.81)	<0.001
	IgG4	0.020	0.245	1.41 (0.99, 1.99)	0.052
	IgG4/IgE ratio	0.009	0.001	0.49 (0.23, 1.04)	0.064
Cockroach	IgE	19.9	33.7	1.21 (1.03, 1.43)	0.024

IgG4	0.065	0.405	1.13 (0.85, 1.50)	0.395
IgG4/IgE ratio	0.004	0.013	0.71 (0.48, 1.04)	0.075
Total IgE	1533	2802	1.24 (1.03, 1.49)	0.025
Total IgG4	89885	156204	1.09 (0.86, 1.38)	0.438
Total IgG4/ total IgE ratio	57.43	55.45	0.89 (0.72, 1.11)	0.292
Total IgE/ cockroach IgE ratio	6.41	8.11	1.01 (0.82, 1.26)	0.893
Total IgE/ dust mite IgE ratio	0.937	0.517	0.81 (0.74, 0.89)	<0.001

	<i>No wheeze</i>	<i>Wheeze</i>		
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SWA	IgE	1672	2627	1.15 (0.81, 1.64)	0.425
	IgG4	12753	14802	1.05 (0.71, 1.54)	0.808
SEA	IgE	1636	1547	0.93 (0.66, 1.29)	0.662
	IgG4	81698	145978	1.16 (0.77, 1.73)	0.449
House dust mite	IgE	2.8	26.6	1.35 (0.89, 2.04)	0.148
	IgG4	0.007	0.008	0.85 (0.59, 1.25)	0.407
	IgG4/IgE ratio	0.011	0.001	0.45 (0.22, 0.93)	0.032
Cockroach	IgE	21.2	2.9	0.69 (0.55, 0.87)	0.003
	IgG4	0.021	0.006	0.65 (0.53, 0.82)	0.001
	IgG4/IgE ratio	0.004	0.002	1.14 (0.47, 2.74)	0.760
	Total IgE	1630	1522	0.88 (0.72, 1.07)	0.205
	Total IgG4	100983	74475	0.85 (0.61, 1.17)	0.302
	Total IgG4/ total IgE ratio	61.00	47.36	0.95 (0.77, 1.18)	0.683
	Total IgE/ cockroach IgE ratio	6.29	6.86	0.91 (0.78, 1.05)	0.187

Total IgE/ dust mite IgE ratio	0.857	0.752	0.91 (0.77, 1.05)	0.188
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[†]All antibody concentrations in ng/ml.

[‡]All geometric mean ratios and 95% confidence intervals adjusted for survey design, age and sex.

[¶]Geometric mean ratios and 95% confidence intervals for associations between antibody levels and SPT reactivity and wheeze were additionally adjusted for SmKK result.

SWA: *Schistosoma* worm antigen; **SEA:** *Schistosoma* egg antigen; **SmKK-**: Kato-Katz negative result (*S. mansoni*), single stool sample; **SmKK⁺**: Kato-Katz positive result, single stool sample; **aGMR:** adjusted geometric mean ratio; **95% CI:** 95% confidence interval.

Figure 1. Hypothesised contribution of *S. mansoni*-induced cytokines and antibodies to allergy-related outcomes

